

RESEARCH PAPER

Extended grain filling has potential to improve yield in grain sorghum

Daniel Otwani^{1,*}, Greg McLean², Graeme Hammer², Alan Cruickshank³, Colleen Hunt^{1,3}, Yongfu Tao⁴, Anna Koltunow², Emma Mace^{1,3}, and David Jordan¹

¹ Queensland Alliance for Agriculture and Food Innovation (QAAFI), The University of Queensland, Hermitage Research Facility, Warwick, 4370 QLD, Australia

² Queensland Alliance for Agriculture and Food Innovation (QAAFI), The University of Queensland, St Lucia, 4067 QLD, Australia

³ Agri-Science Queensland, Department of Agriculture and Fisheries (DAF), Hermitage Research Facility, Warwick, 4370 QLD, Australia

⁴ Agricultural Genomics Institute at Shenzhen, Chinese Academy of Agricultural Sciences, Shenzhen, Guangdong 518120, China

* Correspondence: d.otwani@uq.edu.au

Received 8 September 2024; Editorial decision 4 March 2025; Accepted 11 March 2025

Editor: Fernanda Gonzalez, INTA-CONICET, Argentina

Abstract

Yield increase in sorghum has been achieved primarily by increasing grain number. Scope exists to increase yield by increasing grain size; however, this has been limited by the negative correlation between grain size and grain number. Extending the duration of the grain-filling period has potential to enable increased grain size without the trade-off with grain number. This study explored grain-filling duration (GFD) in a diverse panel of 904 sorghum genotypes in three environments across 2 years. Significant variation in GFD observed, ranging from 400 to 680 degree-days, included entries with significantly longer GFD than current commercial hybrids. Longer GFD was shown to result in larger grain size. Additionally, only low associations between GFD and grain number per panicle, flowering time, or plant height were observed, indicating that GFD could be manipulated without adverse penalty to these traits. A simulation study to estimate the benefit of an increased GFD across Australian sorghum-growing environments over 60 years revealed positive impacts on yield when GFD was increased by either 10% or 20% in environments with low and mild post-anthesis water stress but not in environments with sustained terminal water stress. However, maintaining overall crop duration by shortening time to flowering while extending GFD led to neutral or negative effects on yield. These results reveal opportunities to exploit GFD for improved genetic gains for yield in sorghum, especially in environments or seasons where water does not become more limiting post-anthesis.

Keywords: Crop simulation modelling, degree-days, physiological maturity.

Abbreviations: GFD, grain-filling duration; APSIM, Agricultural Production Systems sIMulator; G×E×M, genetics by environment by management.

© The Author(s) 2025. Published by Oxford University Press on behalf of the Society for Experimental Biology. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (<https://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact reprints@oup.com for reprints and translation rights for reprints. All other permissions can be obtained through our RightsLink service via the Permissions link on the article page on our site—for further information please contact journals.permissions@oup.com.

Introduction

Productivity increases in major cereal crops have been attained mainly through increases in grain number per unit area (Peltonen-Sainio *et al.*, 2007; Boyles *et al.*, 2016), as observed in rice and wheat in the green revolution era (Khush, 1999; Reynolds *et al.*, 1999). While grain size also contributes to potential yield, a compensating trade-off between grain size and grain number (Sadras, 2007; Boyles *et al.*, 2016; Khan *et al.*, 2022), together with a limited genetic variation for grain size in cereals, has limited its exploitation for yield improvement. Grain-filling duration (GFD), which is the time period from flowering to the formation of the abscission layer on the grain, and the rate of grain filling, which is the amount of assimilates partitioned to the grain per unit of time, both contribute to the final grain size in crops (Egli, 2006; Gambin *et al.*, 2008; Sadras and Egli, 2008; Xie *et al.*, 2015). A recent study on commercial sorghum hybrids reported no changes in GFD, filling rate, and grain size over the last six decades of breeding in the USA (Demarco *et al.*, 2023). In contrast, kernel weight has made a significant contribution to the genetic gains in maize hybrid yield in the same region, predominantly due to an extended kernel-filling duration in new maize hybrids relative to old hybrids (Fernández *et al.*, 2022). Similar observations have been reported in rice (Yang *et al.*, 2008) and wheat (Chapman *et al.*, 2021), and a preliminary detailed study in sorghum (Yang *et al.*, 2010) suggested that opportunities exist to explore available variation in GFD for yield in breeding programmes (Sadras and Egli, 2008; Gambin and Borrás, 2012).

The interplay between grain-filling rate and duration and their contribution to yield in sorghum have been studied, with inconclusive results (Woldesemayat *et al.*, 2015; Demarco *et al.*, 2023). There have been limited studies on the physiological and genetic control of GFD in sorghum. Yang *et al.* (2010) reported a detailed study on kernel growth in a limited set of sorghum germplasm varying in grain size. They found that the long kernel-filling duration of the large-seeded KS115-based germplasm (Tuinstra *et al.*, 2001) was the main mechanism associated with its increased grain yield. Variability exists in the rate of grain filling in cereals and legumes; however, this has been mostly yield neutral (Egli, 2006, 2017) as a consequence of the trade-offs with grain number. Since potential seed size in sorghum is more limited by genetic potential rather than assimilate availability (Tao *et al.*, 2017, 2021), we hypothesize that with non-limiting assimilates, accounting for environmental variations in temperature, longer grain filling genotypes could provide yield advantage by enabling more assimilates partitioned to the grain (Eastin *et al.*, 1971). Understanding the physiological and genetic control of the nexus of grain traits that contribute to a yield advantage in sorghum could provide new avenues to improve and sustain yields in an increasingly unpredictable production environment. Here we aim to investigate the variation in GFD of diverse sorghum lines across multiple environments and simulate the impact on sorghum yield and yield components of extending the GFD.

Determining the potential value of variation in target traits to breeders and ultimately to producers is difficult without many years of empirical testing. Crop growth modelling (Kiniry and Bockholt, 1998; Hammer *et al.*, 2010) has been used for simulations of complex adaptive traits across target environments to understand the potential importance to breeding and consider their possible value against resource investment (Kholová *et al.*, 2014; Hammer, 2020). The APSIM (Agricultural Production Systems sIMulator) model has been used to model photosynthesis and other traits in sorghum along with agronomic interventions to assess possible benefits to growers in developed and developing regions of the world (Dimes *et al.*, 2003; Wu *et al.*, 2019). Modelling sorghum with an extended grain-filling period remains an open question in research and the potential benefits/losses have not been quantified.

Hence, the objectives of this study were to (i) establish the extent of genetic variation for GFD in a diverse set of sorghum genotypes; (ii) establish the association of GFD with other yield determinants in sorghum; and (iii) examine the putative value of an extended GFD to sorghum yield across environments through simulation modelling.

Materials and methods

Plant materials and experiments

The sorghum diversity panel (DP) ($n=904$) previously described by Tao *et al.* (2020) was used in the current study. Three experiments were planted, two at the Hermitage Research Facility (HRF), Warwick, Queensland, Australia (28°12'S, 152°5'E, 470 m above sea level) in November 2020 and December 2021. A total of 881 DP genotypes were planted in a row-column design with partial replication where 30% of the genotypes were replicated two or more times while the remaining 70% were in single plots in the 2020/21 season (HRF1) and a fully replicated trial in the 2021/22 season (HRF2). The third experiment was planted at Gatton Research Facility (GAT), Gatton, Queensland, Australia, (27°33'S, 152°20'E, 94 m above sea level) in February 2021 (Supplementary Fig. S1). A total of 609 DP genotypes were planted in a fully replicated trial of two replications in a row-column design. All the trials were planted during the Australian summer growing season in single row plots 4 m long with 0.76 m spacing between rows, using an ALMACO GPS-guided and spaced vacuum planter to achieve a planting density of 80 000 plants ha⁻¹. Fertilizer was applied at the rate of 150 kg nitrogen ha⁻¹. Supplemental irrigation was provided when required to avoid water stress. Standard agronomic practices were employed in the trial management to ensure timely pest and weed control. Overall, the experiments had 598 genotypes in common (Table 1). The DP lines were classified into four racial groups based on population structure analysis, as described in Tao *et al.* (2020), as Guinea, Caudatum, Kafir, and Durra (Asian and East African origin), with admixture types designated as mixed.

Table 1. Number of genotypes in each experiment (diagonal) and genotype concurrence across experiments (off-diagonal elements)

Experiment	GAT	HRF1	HRF2
GAT	609		
HRF1	598	881	
HRF2	599	852	879

Phenotypic evaluation

Single plants of each genotype were tagged in each plot at the time of head exsertion prior to onset of flowering (Fig. 1A). All measurements for timing of flowering and maturity were recorded on the tagged plant. Flowering time was recorded as the date when the first anthers become visible at the tip of the panicle (Fig. 1B). The tagged plant was monitored throughout the season, and the date of physiological maturity was recorded as the date when a sampled grain from the tip of the panicle first showed the abscission layer (black layer) at the point of connection of the grain (Fig. 1D, far right grain image). Plant height was measured at HRF2 by selecting one plant at random from the plot and measuring the distance from the base of the plant to the tip of the panicle at physiological maturity. Single panicles were harvested at HRF2, threshed, and cleaned before grains per panicle and thousand kernel weight (TKW) were measured using an automatic seed counter and weighing machine (Ball Coleman Gen3 seed counter). Daily weather data were recorded using a portable weather station placed within the trial to record daily maximum and minimum air temperatures for the duration of the experiment.

Overall, the trial at Gatton experienced lower temperatures during anthesis and post-anthesis in the grain-filling period.

Thermal time calculation

The observations for time (d) to flowering and physiological maturity for each entry were used to calculate the thermal time for grain fill duration using the method described in Hammer and Muchow (1994) to compute degree-days ($^{\circ}\text{Cd}$) for each day during the grain-filling period as:

$$\Delta \text{TT} = 0 \text{ for } \text{TT}_b$$

$$\Delta \text{TT} = T - T_b \text{ for } T_b \leq T \leq T_{\text{opt}}$$

$$\Delta \text{TT} = T_{\text{opt}} - T_b \text{ for } T > T_{\text{opt}}$$

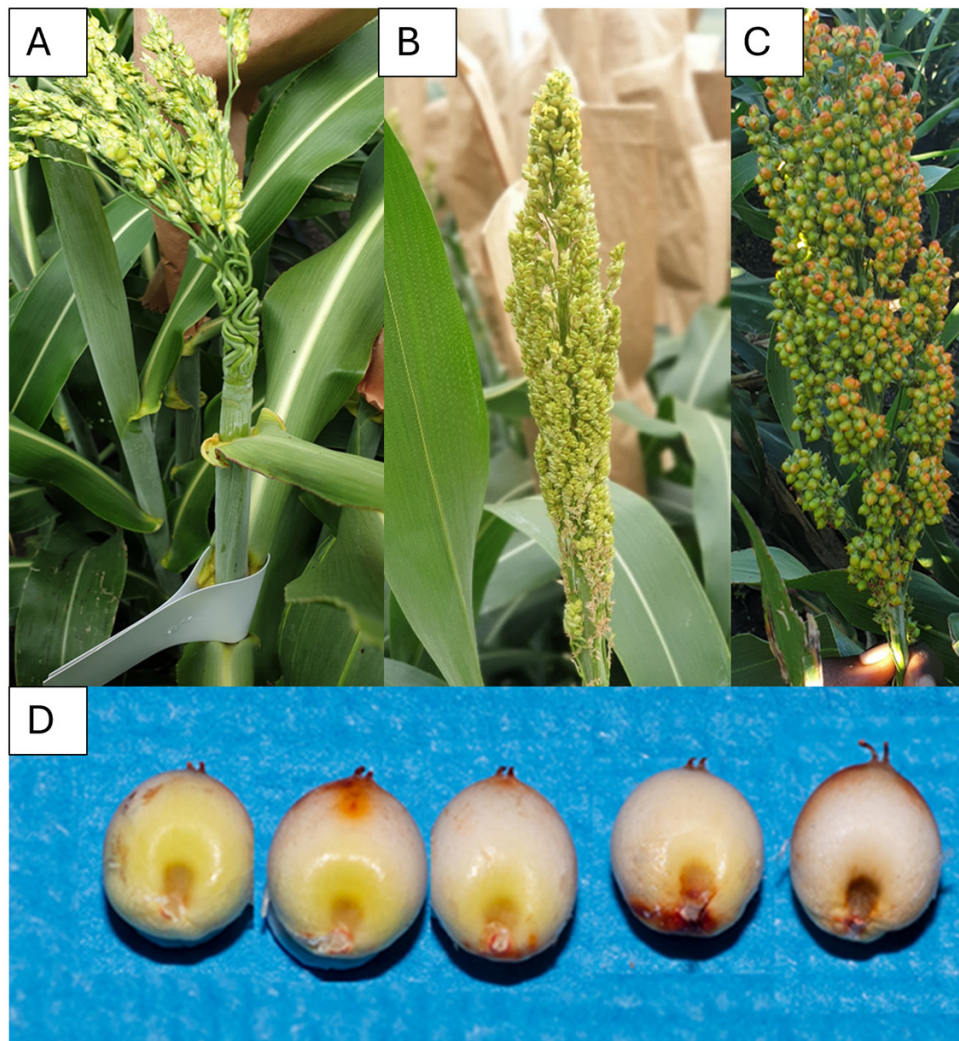


Fig. 1. Images of sorghum plants. (A) Tagged sorghum plant for data measurement. (B) Sorghum head at the onset of flowering when the date of flowering was recorded for the tagged plant. (C) Sorghum head during grain filling, showing how different sides of the sorghum head mature at different rates. (D) Sampled sorghum grains showing progression to formation of the abscission layer (black layer) from left to right; the far right denotes physiological maturity of the grain.

where T_b is base temperature for development ($^{\circ}\text{C}$), T_{opt} is the optimum temperature for development ($^{\circ}\text{C}$), and T is the average daily temperature ($^{\circ}\text{C}$)

The base temperatures for GFD and vegetative growth up to flowering were set at 5.7°C and 11°C , respectively (Hammer and Muchow, 1994), and the optimum temperature for grain filling was taken as 23.5°C (Hammer and Muchow, 1994) and 30°C for vegetative growth. In addition, the maximum temperature for vegetative growth was taken as 42°C (Kumar *et al.*, 2009). Throughout the duration of the experiment, the mean daily temperatures were within the optimum range for all the growth stages. This approach was used because GFD is more associated with accumulated thermal time than with time in days and, given the variation in daily temperature and the variation in flowering dates, it is likely that the use of days to measure GFD would result in over- and underestimation of the GFD for individual lines.

Modelling study

A simulation study was conducted using the APSIM NEXT GEN platform (Holzworth *et al.*, 2018), which incorporates the sorghum crop model detailed by Hammer *et al.* (2019, 2010). Three locations (Supplementary Fig. S1) representing the main sorghum production regions in Australia were used: Emerald in Central Queensland with a black vertosol soil of 1000 mm depth and 160 mm maximum plant-available water content; Dalby in South Queensland with a black vertosol soil of 1800 mm depth and 306 mm maximum plant-available water content; and Tamworth in northern New South Wales with a black vertosol soil of 1800 mm depth and 236 mm maximum plant-available water content as per Hammer *et al.* (2014). Sixty years of historical daily weather data (1960–2020) were used for all the simulations. All simulations were conducted assuming 100 mm available soil water at sowing and non-limiting nitrogen (N) availability. Three sowing dates were simulated for each season: one each in October, November, and December. Planting density was maintained at five plants m^{-2} with a solid row configuration at a row spacing of 1000 mm. Sowing depth was set at 30 mm. Genotypic coefficients were set for the standard commercial hybrid 'Buster' at the values reported by Hammer *et al.* (2010) with tillering as reported by Hammer *et al.* (2014). The output included yield, yield components, leaf area index (LAI) at flowering and maturity, leaf number, fertile tiller number, leaf, stem, and total biomass at flowering and maturity, extractable soil water at flowering and maturity, and days to flowering and maturity. Each simulation was characterized by its environment type (ET1–ET5) depending on the dynamic of the plant water status through the season and its proximity to ETs as described in Hammer *et al.* (2014). Briefly ET1 on average experienced low levels of water limitation throughout the crop season (low stress), ET2 on average experienced an increasing level of water limitation post flowering (mild-stress), ET3 on average experienced an early onset of water limitation that was relieved during the grain-filling period post-anthesis (relieved stress), ET4 on average experienced an early water limitation pre-anthesis that progressed post-anthesis with no relief (sustained stress), and ET5 on average had a gradual water limitation that started pre-flowering and progressed post-flowering (terminal stress).

Three simulation treatments were imposed: (i) standard, where a normal time to flowering and maturity was maintained; (ii) extended, where normal time to flowering was maintained with the flowering to maturity phase being extended by 10% or 20% (~5 d and 10 d, respectively); and (iii) revised, where normal overall crop duration (time to maturity) was maintained but with a reduced time to flowering and grain filling extended by 10% or 20% (~5 d and 10 d, respectively).

Statistical analyses

Data from the three sites were combined in a multi-environment trial (MET) and analysed using a linear mixed model. This model was used to

estimate the correlations between the sites and to calculate heritabilities for each site.

The standard representation of a linear mixed model is given by:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\tau} + \mathbf{Z}\mathbf{u} + \mathbf{e} \quad (1)$$

where $\mathbf{y}$ is the vector of observations with the sites stacked, $\mathbf{X}$ is the design matrix for fixed effects, $\boldsymbol{\tau}$ is the vector of fixed effects, $\mathbf{Z}$ is the design matrix for random effects, $\mathbf{u}$ is the vector of random effects, which has a normal distribution with mean 0 and variance–covariance matrix $\mathbf{G} [\mathbf{u} \sim \mathbf{N}(\mathbf{0}, \mathbf{G})]$, with fixed and random spatial effects included for each site as necessary (Gilmour *et al.*, 1997), and $\mathbf{e}$ is the vector of residuals $\mathbf{e} \sim \mathbf{N}(\mathbf{0}, \mathbf{R})$.

The variance–covariance matrix for the site by genotype interaction (G×E) was fitted using a correlation structure (corgh). This structure allows for a different genetic variance for each site and different correlations for each pair of sites. Different models were fitted separately for each trait, with random and fixed terms included as necessary per site (Supplementary Table S1). The simulation data were analysed using a linear mixed model with yield as response, and simulation type and environment type interaction included as fixed effects. The residual term has a variance–covariance matrix that allowed for separate residual variances per environment type (Supplementary Table S1). All analyses were conducted in R (R Core Team, 2024) environment version 4.04; the package ASReml-R (The VSNi Team, 2023) was used to fit all models, and the package ggplot2 (Wickham, 2016) was used in visualizing all figures.

Results

Climatic conditions

Daily temperatures were recorded at each experimental site (Fig. 2) for the duration of the experiments. At HRF1 and HRF2, average daily temperature throughout the grain-filling period was always above the base temperature for development (5.7°C), so no adjustments were necessary in calculating thermal time. However, mean daily temperature exceeded 23.7°C on a few occasions, and adjustments were made accordingly to account for thermal time accumulated. At GAT, the experiment experienced cooler temperatures around flowering time, with instances where the average daily temperature was below the base temperature, so adjustments were made accordingly. Rainfall was adequate and well distributed throughout the season (data not shown) so that water was not limiting.

Variation in grain-filling duration

Across genotypes, GFD ranged from 20 d to 60 d, with means of 46 (28–60), 30 (22–38), and 34 (20–44) d at GAT, HRF1, and HRF2, respectively. When converted to thermal time, the GFD ranged from 400°Cd to 680°Cd (Fig. 3). The means across the entries for each experiment were 510, 506, and 521°Cd for GAT, HRF1, and HRF2, respectively (Table 2). This indicates the major effect of temperature in generating duration differences across the experiments. The consistency of long GFD genotypes across the three experiments by observing the top 5% of genotypes indicated that >77% were

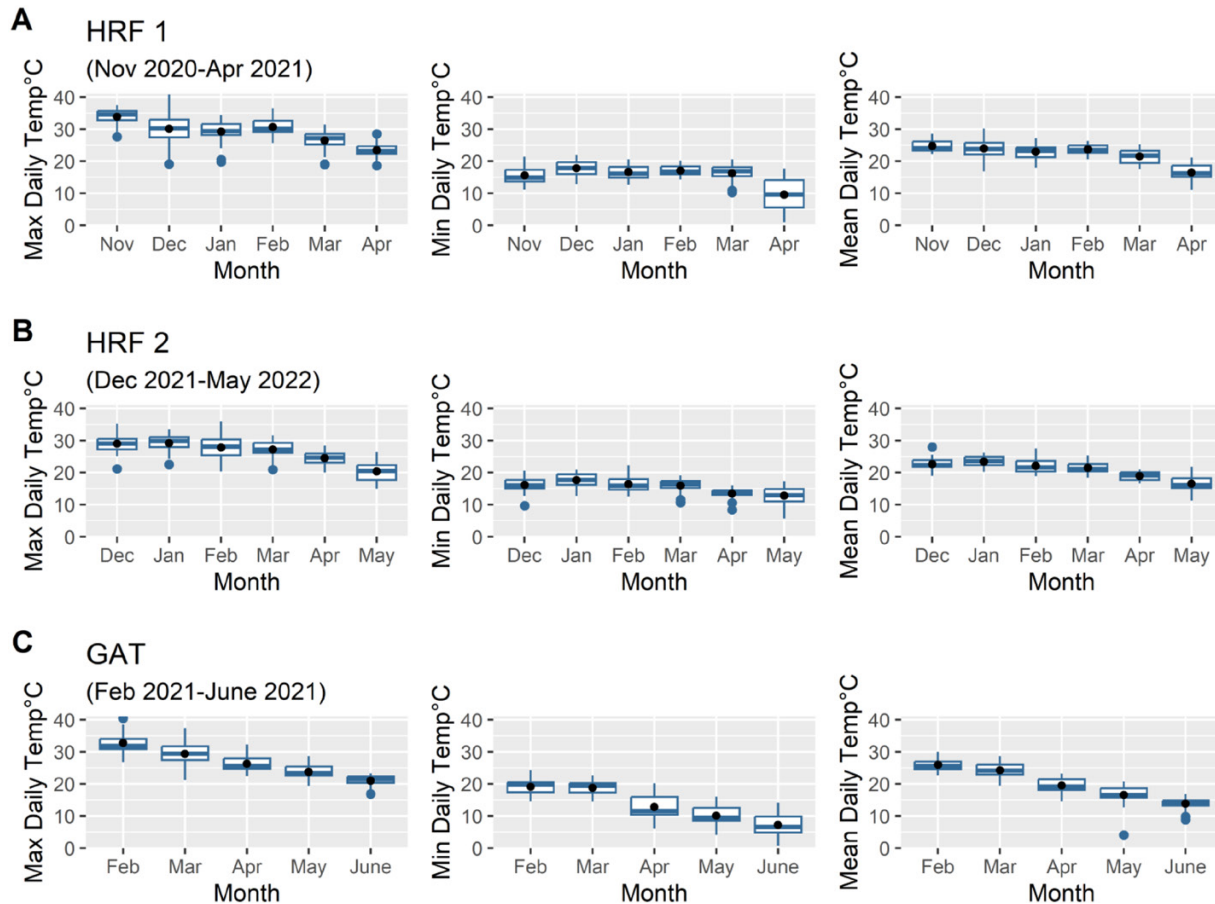


Fig. 2. Maximum, minimum, and mean daily air temperatures. The maximum, minimum, and mean daily temperature from the three field experiments recorded from a portable weather station located within each experiment are shown. HRF1 (A), HRF2 (B), and GAT (C).

consistent (Supplementary Table S2), similar to the observation in the genetic correlations (Table 2).

A comparison of GFD across the sorghum genotypes defined by races showed that race Guinea had a different GFD on average from all the other races (Fig. 4).

Genetic variances, heritability, and correlation of grain-filling duration across experiments

Appreciable genetic variation for GFD was observed, with moderate broad sense heritability estimates ranging between 41% and 61%. Genetic correlations between sites were strengthened or unaltered when GFD was estimated in thermal time rather than in days (Table 2), especially where large temperature variations were observed, namely HRF1 and GAT.

Association of grain filling-duration with yield components

Across experiments, flowering time ranged from 523 °Cd in the earlier flowering genotype to 914 °Cd in the late flowering

types. The overall mean flowering time was 734 °Cd. No significant association was observed between GFD and flowering time at HRF1, while at HRF2 and GAT, GFD had a significant association with flowering time, explaining 9% and 1.5%, respectively, of the observed variation (Fig. 5A). Plant height, when measured from the base of the plant to the tip of the panicle, ranged from 58.31 cm to 217.13 cm with a mean of 101.98 cm at HRF2. Variation in plant height explained between 0.8% and 2.8% of observed variation in GFD across the test locations (Fig. 5B). Grain number per panicle ranged from 572 to 3678, with a mean of 1615 grains. Significant variation in grain number was observed at HRF1 and HRF2 but not at GAT (Fig. 5C). The TKW ranged from 9.47 g to 45.3 4g, with a mean of 22.39g. The commercial check had a TKW of 27.82 g. TKW was significantly positively associated with GFD, explaining between 6% and 25% of the observed variation in GFD across the locations (Fig. 5D).

Simulation of grain-filling duration

The average sorghum yield in the simulation study was 4423 (2316–6663) kg ha⁻¹ at Tamworth, 4231 (2430–5480) kg ha⁻¹

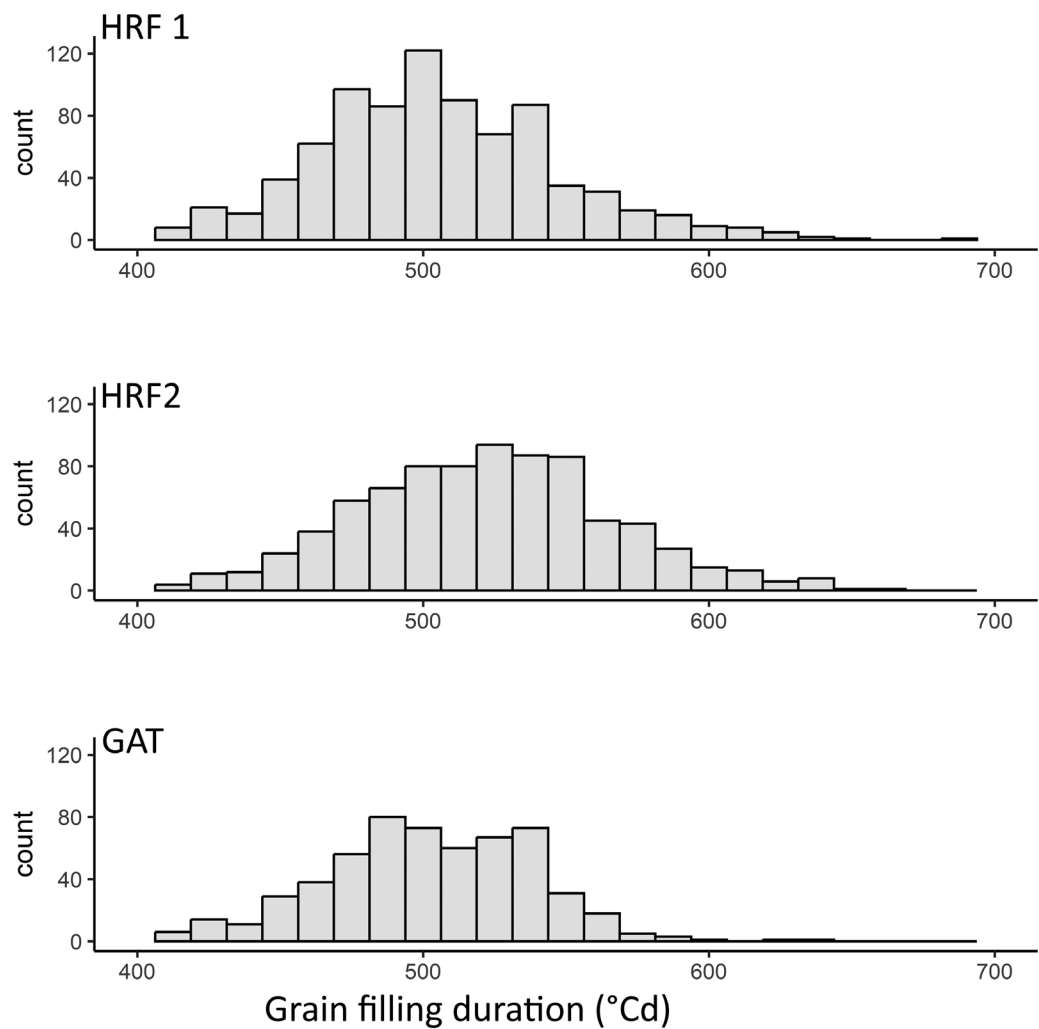


Fig. 3. Histogram showing the distribution of grain-filling duration (GFD) in thermal time for the three experiments at HRF1, HRF2, and GAT, respectively.

Table 2. Mean GFD, genetic variance, and estimate of broad sense heritability (H^2) for each experiment, and genetic correlation of GFD across experiments for measures in days (above diagonal) or degree-days (below diagonal in bold).

	Mean GFD (°Cd)	Genetic variance	Error variance	H^2	Genetic correlations		
					GAT	HRF1	HRF2
GAT	500	757	1250	41	1	0.3	0.46
HRF1	506	1004	1027	61	0.52	1	0.92
HRF2	521	819	1584	56	0.45	0.86	1

at Emerald, and 4438 (2436–7154) kg ha⁻¹ at Dalby. Extending the GFD of the simulated standard hybrid by 10% resulted in increases in yield of between 5.6% and 7.5% relative to the standard hybrid (Fig. 6A; Table 3). A 20% extension of GFD resulted in simulated yield increases of between 10% and 13% (Fig. 6B; Table 3). Reducing the duration to flowering by 10%

or 20% of the standard GFD to enable an extended GFD by either 10% or 20% resulted in both positive and negative changes in yield relative to the standard genotype (Fig. 6C, D). Further evaluation of the simulated sorghum yields across the different environment types within each location yields revealed mean yields ranging from 2436 kg ha⁻¹ in severely water-limited environment type 4 (ET4) to 7154 kg ha⁻¹ in environment type 1 (ET1) where water was mostly not limiting (Table 4). Extending GFD by 10% resulted in a yield increase of between 3.3% and 8.5% across the environment types, with the highest percentage increase observed in ET3. Changes of between 6.4% and 16.6% were observed across the environment types in scenarios where the GFD was extended by 20%. These yield increases were mainly significantly different from the standard in environments where water was not limited post-anthesis (Table 4; Supplementary Table S3). In the revised scenarios where the shorter time to flowering was simulated, there was generally a non-significant change in yields across all the locations and environment types, with few exceptions

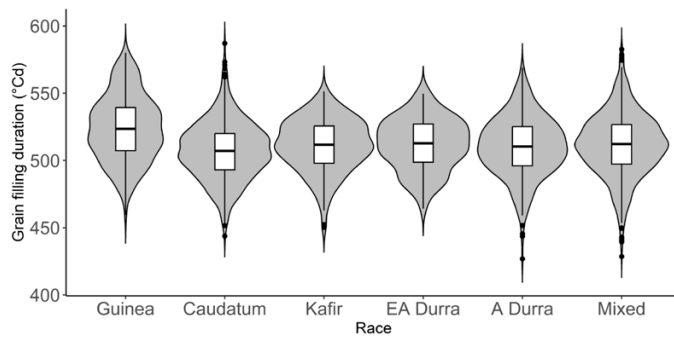


Fig. 4. Grain-filling duration (GFD) in thermal time across the sorghum racial groups (Race) across the three experiments.

at ET2 Dalby, ET1 and ET2 Emerald, and ET4 Tamworth where significant positive and negative changes were observed (Supplementary Table S3). Reducing the flowering time to increase GFD at Tamworth generally had negative impacts on the attainable yield across the different environment type simulations (Supplementary Table S3).

Increases in kernel weight were observed in all the environments and environment types, with greater increases observed in ET1 and ET2, where water was mostly non-limiting post-flowering. ET4 and ET5, which have significant post-flowering water limitation, had lower near zero increases in kernel weight (Supplementary Fig. S2).

Discussion

This study reports the extent of genetic variation in GFD observed in a diverse panel of sorghum genotypes and highlights opportunities to use this trait in breeding for yield. A simulation study was conducted that suggested that increasing GFD has the potential to result in yield increases likely in Australian production environments.

Variation in grain-filling duration in sorghum is available beyond the commercial range but does not appear to have been utilized for improving yield

Appreciable genetic variation with moderate heritability was observed for GFD in the diversity panel, with the phenotypic distribution indicating that the trait is controlled by multiple genes. We observed that 10% of genotypes evaluated have a longer GFD than the commercial check and did not seem to be negatively associated with important agronomic traits such as height and phenology. In combination, our results suggest that there is potential to increase yield by enhancing GFD beyond current levels. However, previous reports on changes in GFD across sorghum hybrids in the USA showed no changes in GFD over 60 years of breeding associated with a 0.5% per year increase in yield over the same period (Demarco *et al.*, 2023). The lack of association of GFD with yield in the study of Demarco

et al. (2023) could be explained in at least four ways: (i) that GFD was poorly estimated; (ii) that GFD is not associated with enhanced yield in sorghum; (iii) that GFD is correlated with other traits that have negative consequences on hybrid performance; and (iv) that there is no genetic variation for GFD that can be exploited in elite US sorghum populations.

It is highly unlikely that GFD was poorly estimated by Demarco *et al.* (2023) given the narrow range of relative maturity in the tested hybrids and the limited difference in temperature across all the test locations. Regarding point (ii), several studies in cereals have reported the contribution of GFD to yield and yield genetic gain. In maize, Fernández *et al.* (2022) reported that GFD contributed close to 50% of the genetic gain in kernel weight. Similar observations have been reported in a historical study of Chinese maize hybrids between 1964 and 2014 reporting that GFD contributed up to 54.46% of the observed variation in hundred kernel weight, with recent hybrids having longer filling duration and yield than their predecessors (Gao *et al.*, 2023). Reports of longer GFD association with yield have also been recorded in wheat (Chapman *et al.*, 2021) and rice (Jones *et al.*, 1979; Wang *et al.*, 2008; Yang *et al.*, 2008). These observations show that GFD is associated with yield in cereals and therefore is likely to be similarly associated in sorghum, as observed by Gizzi and Gambin (2016). Regarding point (iii), in the current study in sorghum, TKW has been shown to have a significant positive association with GFD. Additionally, we have no evidence to indicate that GFD is negatively associated with major agronomic traits such as height and flowering time, though the potential association between increased GFD, resulting in increased stem remobilization, and increased lodging needs to be explored. Finally, regarding point (iv), it is plausible that there is no substantial exploitable genetic variation for GFD in elite US sorghum germplasm potentially due to a small genetic pool that characterizes many sorghum hybrid breeding programmes. Most long GFD genotypes were from the Guinea race (Fig. 4) which is seldom used in commercial hybrid programmes. Sadras and Egli (2008), while reporting that grain-filling rate dominated contribution to yield, appreciated that with wider diversity of genotypes, the contribution of GFD to yield would be captured better. Understanding the GFD trait in sorghum therefore could provide opportunities for increasing genetic gains for grain size and therefore yield.

Simulation modelling indicates the potential of grain-filling duration to increase yield in sorghum

In this study, GFD was modelled by increasing the length of time between flowering and physiological maturity, assuming that the rate of grain filling did not change, and that maximum grain size was constrained only by the assimilate partitioned to the grain during the period of grain filling. It was assumed that there was no association between GFD and grain number, and the study did not consider the potential increase in lodging that would be likely to be associated with an increase in demand for

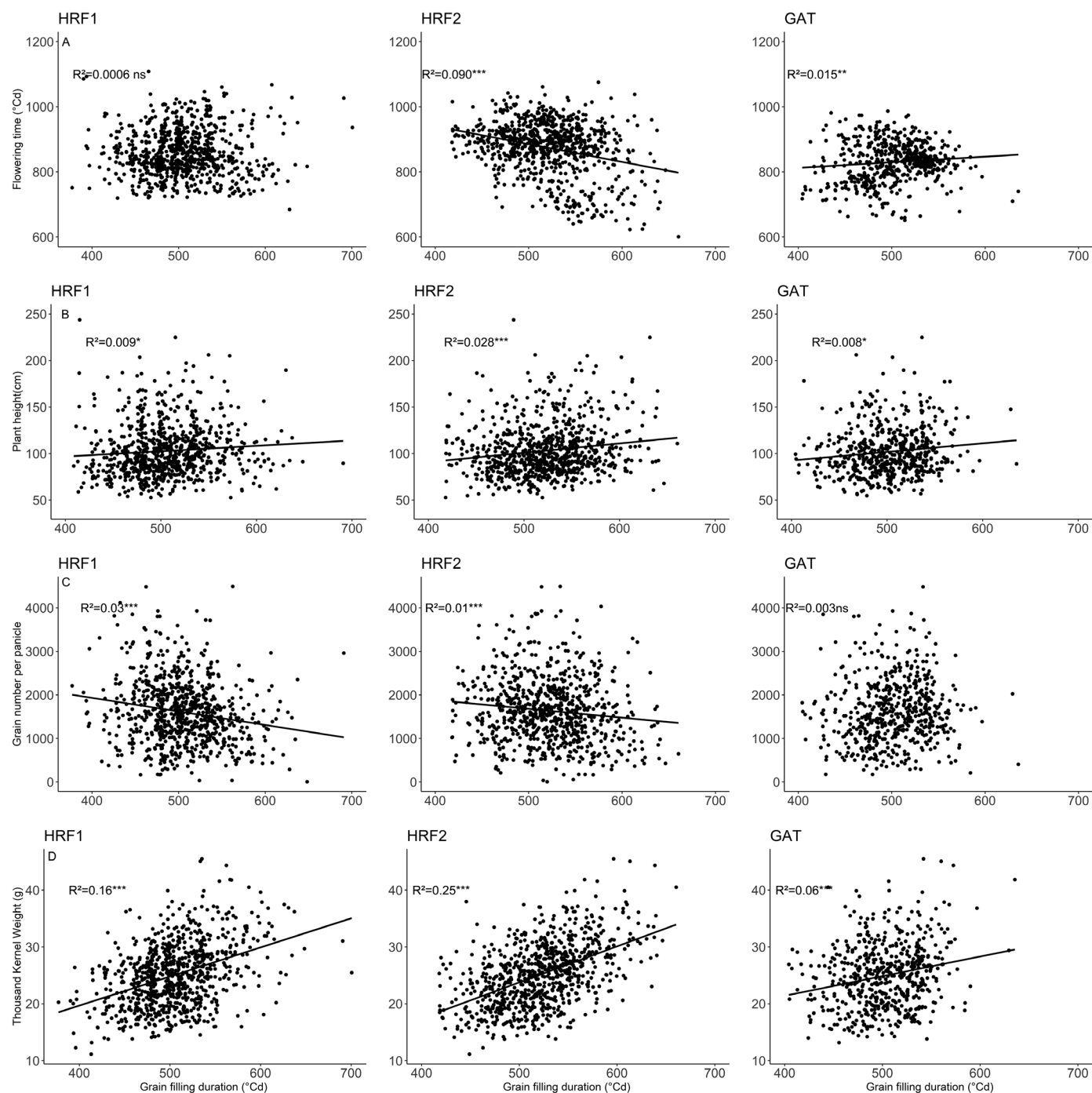


Fig. 5. Scatter plots of sorghum physiological traits versus grain-filling duration for each of the three experiments. (A) Flowering time, (B) plant height, (C) grain number per panicle, and (D) thousand kernel weight (TKW).

carbohydrates. With these assumptions, the simulations suggest that extending the GFD in sorghum could result in increases in grain yield in most Australian sorghum environments. The yield increases were greatest in environments where adequate water was available post-flowering and had the smallest impacts in water-limited environments (Supplementary Table S3). Because of its perennial nature, if water is available during the

grain-filling period, sorghum will continue to produce additional carbohydrate which can be used to increase grain yield.

In Australia, ~50% of the environments in the major sorghum-growing regions have sufficient water available post-flowering to enable extended GFD to result in increased yield. In environments where water is limited, one might expect that the additional demand for assimilate created by a longer GFD

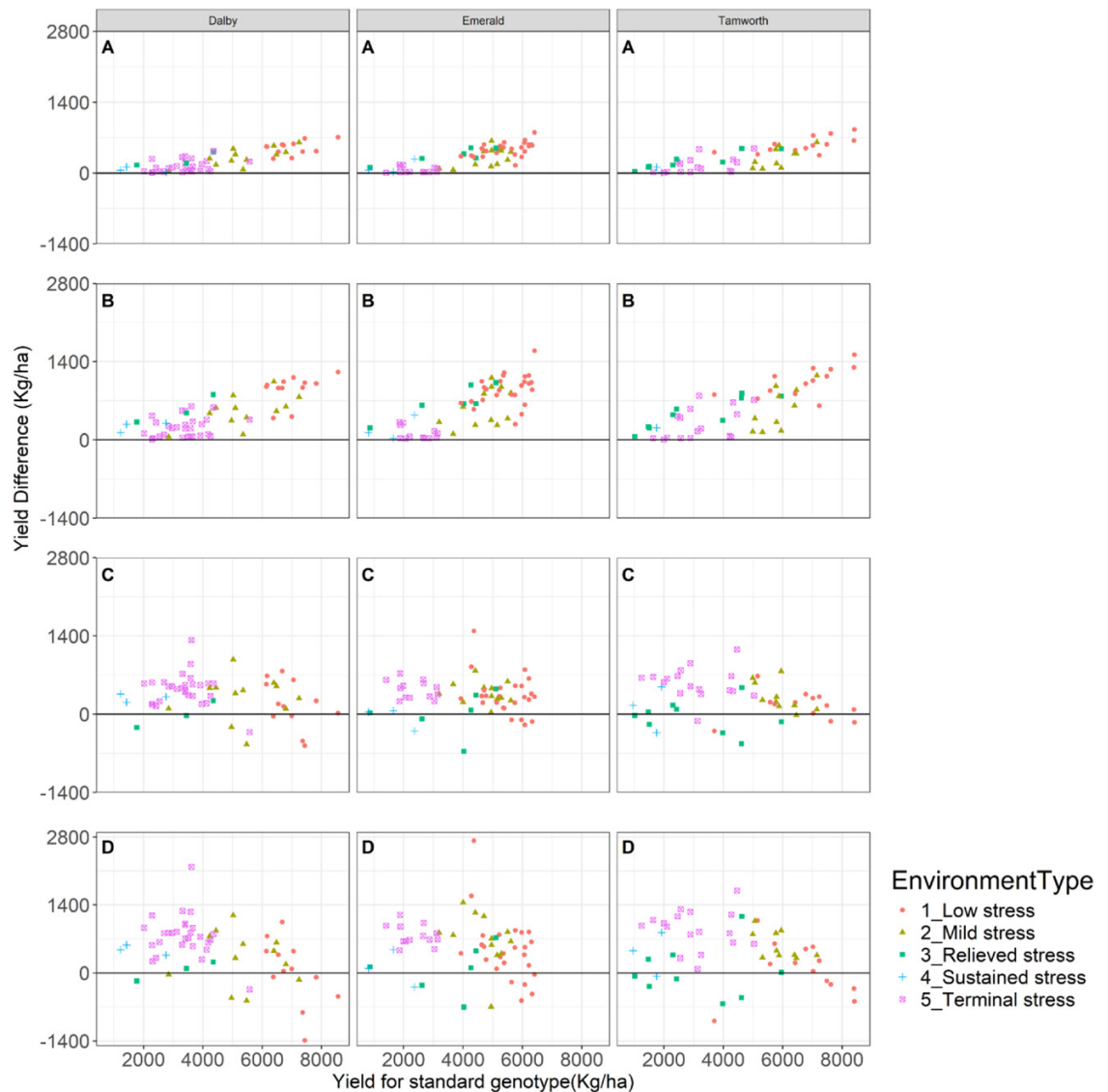


Fig. 6. Simulated yield change versus yield. The simulated yield change versus yield for the standard hybrid for each of 60 years (1960–2020) across the three locations (Dalby, Emerald, and Tamworth) and environment types (ET). (A) For a 10% increase and (B) for a 20% increase in grain-filling duration, (C) for a 10% and (D) for a 20% increase in grain-filling duration with a shorter time to flowering.

would result in remobilization stress, which could increase the degree of lodging. It is likely that traits that achieve temporal water conservation, such as stay green traits (Borrell *et al.*, 2001; Harris *et al.*, 2007; Jordan *et al.*, 2012), and canopy characteristics (Borrell *et al.*, 2014; Mantilla-Perez *et al.*, 2020) that favour increased storage of assimilates such as plant height (Fernandez *et al.*, 2009; George-Jaeggli *et al.*, 2011, 2021), could interact favourably with GFD to enhance its positive impacts on yield.

Different G×E×M strategies for deploying the grain-filling duration trait in sorghum breeding

All simulations conducted revealed that extending the GFD in sorghum provided opportunities for increased or stable yield

in the Australian production environments. The yield increases were consistent with the available water post-anthesis, with minimal and negative increases where sustained water limitation was more severe. The genetic variation in stay green traits (Borrell *et al.*, 2001; Harris *et al.*, 2007; Jordan *et al.*, 2012), plant height (Fernandez *et al.*, 2009; George-Jaeggli *et al.*, 2011, 2021), and canopy characteristics (Borrell *et al.*, 2014; Mantilla-Perez *et al.*, 2020) that can be employed to achieve temporal water conservation in sorghum could be leveraged along with an extended GFD to improve sorghum yield. These strategies could be further tested by simulation modelling. Agronomic interventions, such as planting density (Wade and Douglas, 1990; Whish *et al.*, 2005) and time of planting, could also be important as a strategy to minimize risks of post-anthesis water

Table 3. Simulated percentage change in yield relative to the standard hybrid for genotypes with 10% or 20% longer grain-filling duration

Site	Standard mean yield kg ha ⁻¹	GFD+10% change in yield (%)	GFD+20% change in yield (%)
Tamworth	4423	5.7*	10.5*
Emerald	4231	7.5*	13.7*
Dalby	4438	5.6*	10.1*

The mean of all environments for each site has been used. *Denotes significant difference at $P < 0.05$.

Table 4. Simulated percentage change in yield relative to the standard hybrid for genotypes with 10% or 20% longer grain-filling duration for each environment type (ET) at Dalby

Dalby	Standard mean yield kg ha ⁻¹	GFD+10% change in yield (%)	GFD+20% change in yield (%)
ET1	7154	7.7*	14.5*
ET2	6050	6.2ns	10*
ET3	3070	8.5ns	16.6ns
ET4	2436	4.4ns	9.2ns
ET5	3519	3.3ns	6.4ns

* Denotes significant difference at $P < 0.05$; ns=not significant.

stress. They can also be assessed initially using simulation of $G \times M \times E$ combinations as in the study of [Hammer *et al.* \(2014\)](#).

Reducing the time to flowering to achieve an extended GFD by either 10% or 20% showed increases in yield predominantly in environments where water was limiting (ET4 and ET5), and opposite effects in environments where water was not severely limited post-anthesis (ET1 and ET2) ([Fig. 6](#)). These observations could primarily be due to the fact that (i) the early flowering scenarios provided an opportunity to conserve available moisture which was used during the grain-filling period, increasing the potential yields attained and (ii) the early flowering in scenarios where water was not severely limited resulted in less biomass accumulation pre-flowering and led to a penalty in yield attainable potentially from a reduction in grain numbers ([Van Oosterom and Hammer, 2008](#)).

The Australian production environments simulated here are mostly constrained by the timing and magnitude of water limitation, rather than by season length ([Hammer and Muchow, 1994](#)). In other production regions that might be constrained in length by low temperatures and frost, the extended GFD may have more downside consequences. Again, potential consequences could be explored by simulation in the first instance. In that situation, other strategies, such as maintaining overall crop duration but with extended GFD and slightly earlier flowering, might be advantageous in utilizing the variation in GFD. A potential negative consequence of increasing GFD for

yield advantage in sorghum could occur if the yield increases predispose the crop to lodging. Lodging could occur either because of the heavy panicles attained or due to weaker stems following excessive remobilization of assimilates into the panicles late in the season ([Rajewski and Francis, 1991](#)). The potential impact of excess remobilization has not been captured in the current simulation and would improve future studies of extending GFD. Finally, changes in the GFD may require changes in crop husbandry practices such as fertilizer application, planning of rotations, and spray out activities before sorghum harvest. These should be considered with respect to resources needed and farm profitability.

Potential limitations of the study

Two methodological issues should be noted. First, the estimation of GFD in the current study was based on the observation of the abscission layer (black layer). This allowed at-scale phenotyping to capture the genetic variation for this trait at the population level. In sorghum, ([Vanderlip and Reeves, 1972](#); [Eastin *et al.*, 1973](#)), as well as in maize ([Hunter *et al.*, 1991](#)), the observation of the black layer is a confirmation that the plant reached physiological maturity, but the actual maximum dry weight may occur a few days earlier. Then our GFD based on the black layer may be slightly overestimated. However, the second issue may be underestimating the duration of GFD. We based the estimation on grains from the tip of the panicle. Physiological maturity is observed typically at the base of the panicle at the plant level in sorghum, but tracking GFD from the basal grains presents a complexity in terms of estimating flowering times at scale. While intra-panicle patterns of growth and development differ among grain positions ([Heiniger *et al.*, 1993a](#); [Gambín and Borrás, 2005](#)), sampling consistently from a specified portion of the panicle captures the estimates of GFD at scale appropriately, as observed in a variable pollination study ([Heiniger *et al.*, 1993b](#)). Consequently, estimates of GFD from the top of the panicle may not be necessarily representative of the base of the panicle due to flowering time and genotypic differences, and warrants further investigation.

Conclusion

We have established that exploitable genetic variation is available for GFD in sorghum, with some exotic lines having longer GFD than current commercial hybrids. Simulation studies revealed that extending the GFD increased yield in simulated scenarios in Australian production environments with non-limiting water post-anthesis. While there are factors that require further study, we consider that GFD is a potentially useful target trait for increasing sorghum yields, and further work is needed to understand its genetic control and interactions with crop management.

Supplementary data

The following supplementary data are available at [JXB online](#).

Table S1. The model parameters specified for each of the environments and the simulation experiment.

Table S2. A comparison of concurrence of the top 5% genotypes with extended grain-filling duration across the three sites.

Table S3. Simulated mean yields and standard errors across three locations and the five environment types.

Table S4. Frequency of occurrence of the different environment types across the three locations based on weather data (rainfall) over 60 years (1960–2020).

Fig. S1. A map of Australia showing the location of experiments planted (with green points) and locations simulated in the crop modelling platform APSIM (with blue points). Map modified from Google Maps on 20 December 2024.

Fig. S2. Scatter plot of simulated change in kernel weight and base kernel weight when grain-filling duration was increased by 10%.

Acknowledgements

The authors acknowledge the contribution of the University of Queensland and the Queensland Government's sorghum pre-breeding field team.

Author contributions

DJ, DO, and EM: conceptualization; DO and CH: data curation, formal analysis, and validation; DJ: funding acquisition; DO: investigation, methodology, visualization, and writing—original draft; DJ and EM: project administration; DJ, EM, and AC: resources; DO, CH, GH, and GM: software; AK, EM, AC, CH, and DJ: supervision; DO, DJ, EM, AK, YT, GH, CH, and GM: writing—review & editing.

Conflict of interest

No conflict of interest declared.

Funding statement

The study was funded from investments by the Queensland Government, and The University of Queensland. DO is a beneficiary of the University of Queensland RTP Scholarship.

Data availability

All primary data to support the findings of this study are openly available in Dryad at <https://doi.org/10.5061/dryad.hhmgqnkrn> (Otwani *et al.*, 2025).

References

Borrell A, Hammer G, Van Oosterom E. 2001. Stay-green: a consequence of the balance between supply and demand for nitrogen during grain filling? *Annals of Applied Biology* **138**, 91–95.

Borrell AK, Mullet JE, George-Jaeggli B, van Oosterom EJ, Hammer GL, Klein PE, Jordan DR. 2014. Drought adaptation of stay-green sorghum is associated with canopy development, leaf anatomy, root growth, and water uptake. *Journal of Experimental Botany* **65**, 6251–6263.

Boyles RE, Cooper EA, Myers MT, Brenton Z, Rauh BL, Morris GP, Kresovich S. 2016. Genome-wide association studies of grain yield components in diverse sorghum germplasm. *The Plant Genome* **9**, doi: [10.3835/plantgenome2015.09.0091](https://doi.org/10.3835/plantgenome2015.09.0091).

Chapman EA, Orford S, Lage J, Griffiths S. 2021. Delaying or delivering: identification of novel NAM-1 alleles that delay senescence to extend wheat grain fill duration. *Journal of Experimental Botany* **72**, 7710–7728.

Demarco PA, Mayor L, Rotundo JL, Prasad PVV, Morris GP, Fernandez JA, Tamagno S, Hammer G, Messina CD, Ciampitti IA. 2023. Retrospective study in U.S. commercial sorghum breeding: II. Physiological changes associated to yield gain. *Crop Science* **63**, 867–878.

Dimes J, Twomlow S, Carberry P. 2003. Application of APSIM in small holder farming systems in the semi-arid tropics. In: Struif Bontkes TE, Wopereis MCS, eds. *Decision support tools for smallholder agriculture in sub-Saharan Africa: a practical guide*. Muscle Shoals, AL and Wageningen, The Netherlands: IFDC and CTA, 85–99.

Eastin J, Sullivan C, Ross W, Clegg M, Maranville J. 1971. Comparative development stages in sorghum hybrids and parent lines. In: *Research in the physiology of yield and management of sorghum in relation to genetic improvement*. Co-operative Research by the University of Nebraska Crops Research Division, ARS, USDA and the Rockefeller Foundation, Annual Report No. 5, 168–181.

Eastin JD, Hultquist JH, Sullivan CY. 1973. Physiologic maturity in grain sorghum 1. *Crop Science* **13**, 175–178.

Egli D. 2006. The role of seed in the determination of yield of grain crops. *Australian Journal of Agricultural Research* **57**, 1237–1247.

Egli D. 2017. Seed growth rate and seed-fill duration: variation and regulation. In: *Seed biology and yield of grain crops*, Vol. 2. Wallingford, UK: CABI Publishing, 42–76.

Fernández JA, Messina CD, Salinas A, Prasad PV, Nippert JB, Ciampitti IA. 2022. Kernel weight contribution to yield genetic gain of maize: a global review and US case studies. *Journal of Experimental Botany* **73**, 3597–3609.

Fernandez MGS, Becraft PW, Yin Y, Lübberstedt T. 2009. From dwarves to giants? Plant height manipulation for biomass yield. *Trends in Plant Science* **14**, 454–461.

Gambín BL, Borrás L. 2005. Sorghum kernel weight. *Crop Science* **45**, 553–561.

Gambín BL, Borrás L. 2012. Genotypic diversity in sorghum inbred lines for grain-filling patterns and other related agronomic traits. *Crop and Pasture Science* **62**, 1026–1036.

Gambín BL, Borrás L, Otegui ME. 2008. Kernel weight dependence upon plant growth at different grain-filling stages in maize and sorghum. *Australian Journal of Agricultural Research* **59**, 280–290.

Gao X, Li Y-X, Yang M-T, Li C-H, Song Y-C, Wang T-Y, Li Y, Shi Y-S. 2023. Changes in grain-filling characteristics of single-cross maize hybrids released in China from 1964 to 2014. *Journal of Integrative Agriculture* **22**, 691–700.

George-Jaeggli B, Jordan DR, van Oosterom EJ, Hammer GL. 2011. Decrease in sorghum grain yield due to the *dw3* dwarfing gene is caused by reduction in shoot biomass. *Field Crops Research* **124**, 231–239.

George-Jaeggli B, Lefèvre-Arbogast S, Hunt C, Cruickshank A, Jordan DR. 2021. Tall 3-dwarfs: oxymoron or opportunity to increase grain yield in sorghum? *Planta* **253**, 110.

Gilmour AR, Cullis BR, Verbyla AP, Verbyla AP. 1997. Accounting for natural and extraneous variation in the analysis of field experiments. *Journal of Agricultural Biological and Environmental Statistics* **2**, 269–293.

Gizzi G, Gambin BL. 2016. Eco-physiological changes in sorghum hybrids released in Argentina over the last 30 years. *Field Crops Research* **188**, 41–49.

- Hammer G.** 2020. The roles of credibility and transdisciplinarity in modelling to support future crop improvement. In *Silico Plants* **2**, diaa004.
- Hammer G, Messina C, Wu A, Cooper M.** 2019. Biological reality and parsimony in crop models—why we need both in crop improvement! In *Silico Plants* **1**, diz010.
- Hammer GL, McLean G, Chapman S, Zheng B, Doherty A, Harrison MT, van Oosterom E, Jordan D.** 2014. Crop design for specific adaptation in variable dryland production environments. *Crop and Pasture Science* **65**, 614–626.
- Hammer GL, Muchow RC.** 1994. Assessing climatic risk to sorghum production in water-limited subtropical environments I. Development and testing of a simulation model. *Field Crops Research* **36**, 221–234.
- Hammer GL, van Oosterom E, McLean G, Chapman SC, Broad I, Harland P, Muchow RC.** 2010. Adapting APSIM to model the physiology and genetics of complex adaptive traits in field crops. *Journal of Experimental Botany* **61**, 2185–2202.
- Harris K, Subudhi P, Borrell A, Jordan D, Rosenow D, Nguyen H, Klein P, Klein R, Mullet J.** 2007. Sorghum stay-green QTL individually reduce post-flowering drought-induced leaf senescence. *Journal of Experimental Botany* **58**, 327–338.
- Heiniger RW, Vanderlip RL, Kofoid KD.** 1993a. Caryopsis weight patterns within the sorghum panicle. *Crop Science* **33**, 543–549.
- Heiniger RW, Vanderlip RL, Kofoid KD.** 1993b. Influence of pollination pattern on intrapanicle caryopsis weight in sorghum. *Crop Science* **33**, 549–555.
- Holzworth D, Huth NI, Fainges J, Brown H, Zurcher E, Cichota R, Verrall S, Herrmann NI, Zheng B, Snow V.** 2018. APSIM next generation: overcoming challenges in modernising a farming systems model. *Environmental Modelling & Software* **103**, 43–51.
- Hunter J, TeKrony D, Miles D, Egli D.** 1991. Corn seed maturity indicators and their relationship to uptake of carbon-14 assimilate. *Crop Science* **31**, 1309–1313.
- Jones DB, Peterson ML, Geng S.** 1979. Association between grain filling rate and duration and yield components in rice. *Crop Science* **19**, 641–644.
- Jordan DR, Hunt CH, Cruickshank A, Borrell A, Henzell R.** 2012. The relationship between the stay-green trait and grain yield in elite sorghum hybrids grown in a range of environments. *Crop Science* **52**, 1153–1161.
- Khan N, Zhang Y, Wang J, et al.** 2022. TaGSNE, a WRKY transcription factor, overcomes the trade-off between grain size and grain number in common wheat and is associated with root development. *Journal of Experimental Botany* **73**, 6678–6696.
- Kholová J, Murugesan T, Kaliamoorthy S, et al.** 2014. Modelling the effect of plant water use traits on yield and stay-green expression in sorghum. *Functional Plant Biology* **41**, 1019–1034.
- Khush GS.** 1999. Green revolution: preparing for the 21st century. *Genome* **42**, 646–655.
- Kiniry JR, Bockholt AJ.** 1998. Maize and sorghum simulation in diverse Texas environments. *Agronomy Journal* **90**, 682–687.
- Kumar SR, Hammer GL, Broad I, Harland P, McLean G.** 2009. Modelling environmental effects on phenology and canopy development of diverse sorghum genotypes. *Field Crops Research* **111**, 157–165.
- Mantilla-Perez MB, Bao Y, Tang L, Schnable PS, Salas-Fernandez MG.** 2020. Toward 'smart canopy' sorghum: discovery of the genetic control of leaf angle across layers. *Plant Physiology* **184**, 1927–1940.
- Otwani D, McLean G, Hammer G, Cruickshank A, Hunt C, Tao Y, Mace E, Jordan D.** 2025. Data from: Extended grain filling has potential to improve yield in grain sorghum. Dryad Depository <https://doi.org/10.5061/dryad.hmgqgnkm>
- Peltonen-Sainio P, Kangas A, Salo Y, Jauhiainen L.** 2007. Grain number dominates grain weight in temperate cereal yield determination: evidence based on 30 years of multi-location trials. *Field Crops Research* **100**, 179–188.
- Rajewski JF, Francis CA.** 1991. Defoliation effects on grain fill, stalk rot, and lodging of grain sorghum. *Crop Science* **31**, 353–359.
- R Core Team.** 2024. R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.
- Reynolds M, Rajaram S, Sayre K.** 1999. Physiological and genetic changes of irrigated wheat in the post-green revolution period and approaches for meeting projected global demand. *Crop Science* **39**, 1611–1621.
- Sadras VO.** 2007. Evolutionary aspects of the trade-off between seed size and number in crops. *Field Crops Research* **100**, 125–138.
- Sadras VO, Egli D.** 2008. Seed size variation in grain crops: allometric relationships between rate and duration of seed growth. *Crop Science* **48**, 408–416.
- Tao Y, Mace ES, Tai S, Cruickshank A, Campbell BC, Zhao X, Van Oosterom EJ, Godwin ID, Botella JR, Jordan DR.** 2017. Whole-genome analysis of candidate genes associated with seed size and weight in sorghum bicolor reveals signatures of artificial selection and insights into parallel domestication in cereal crops. *Frontiers in Plant Science* **8**, 1237.
- Tao Y, Trusov Y, Zhao X, et al.** 2021. Manipulating assimilate availability provides insight into the genes controlling grain size in sorghum. *The Plant Journal* **108**, 231–243.
- Tao Y, Zhao X, Wang X, Hathorn A, Hunt C, Cruickshank AW, van Oosterom EJ, Godwin ID, Mace ES, Jordan DR.** 2020. Large-scale GWAS in sorghum reveals common genetic control of grain size among cereals. *Plant Biotechnology Journal* **18**, 1093–1105.
- The VSNi Team.** 2023. asreml: Fits Linear Mixed Models using REML. R package version 4.2.0.302. <https://asreml.kb.vsnr.co.uk/>
- Tuinstra M, Liang G, Hicks C, Kofoid K, Vanderlip R.** 2001. Registration of KS 115 sorghum. *Crop Science* **41**, 932–933.
- Vanderlip RL, Reeves HE.** 1972. Growth stages of sorghum [*Sorghum bicolor*, (L.) Moench]. *Agronomy Journal* **64**, 13–16.
- Van Oosterom E, Hammer G.** 2008. Determination of grain number in sorghum. *Field Crops Research* **108**, 259–268.
- Wade L, Douglas A.** 1990. Effect of plant density on grain yield and yield stability of sorghum hybrids differing in maturity. *Australian Journal of Experimental Agriculture* **30**, 257–264.
- Wang E, Wang J, Zhu X, et al.** 2008. Control of rice grain-filling and yield by a gene with a potential signature of domestication. *Nature Genetics* **40**, 1370–1374.
- Whish J, Butler G, Castor M, Cawthray S, Broad I, Carberry P, Hammer G, McLean G, Routley R, Yeates S.** 2005. Modelling the effects of row configuration on sorghum yield reliability in north-eastern Australia. *Australian Journal of Agricultural Research* **56**, 11–23.
- Wickham H.** 2016. ggplot2: elegant graphics for data analysis. New York: Springer-Verlag.
- Woldesemayat MY, Mekbib F, Gebeyehu S.** 2015. Genetic gain in lowland sorghum [*Sorghum bicolor* (L.) Moench] varieties in Ethiopia. *International Journal of Horticulture and Plant Breeding Sciences* **2**, 1–13.
- Wu A, Hammer GL, Doherty A, von Caemmerer S, Farquhar GD.** 2019. Quantifying impacts of enhancing photosynthesis on crop yield. *Nature Plants* **5**, 380–388.
- Xie Q, Mayes S, Sparkes DL.** 2015. Carpel size, grain filling, and morphology determine individual grain weight in wheat. *Journal of Experimental Botany* **66**, 6715–6730.
- Yang W, Peng S, Dionisio-Sese ML, Laza RC, Visperas RM.** 2008. Grain filling duration, a crucial determinant of genotypic variation of grain yield in field-grown tropical irrigated rice. *Field Crops Research* **105**, 221–227.
- Yang Z, van Oosterom EJ, Jordan DR, Doherty A, Hammer GL.** 2010. Genetic variation in potential kernel size affects kernel growth and yield of sorghum. *Crop Science* **50**, 685–695.